



## **A COMPARATIVE CLINICAL EVALUATION IN NATURAL VERSUS STIMULATED IVF CYCLES EMBRYO**

### **Corresponding Author**

**Dr . Humera Bint Raees**

*Laparoscopic Gynae Surgeon and Specialist Obstetrician*

*Zulekha Hospital*

*Sharjah UAE*

*Email- [drhumera\\_raees@yahoo.com](mailto:drhumera_raees@yahoo.com)*

**INTRODUCTION:** The impact of controlled ovarian stimulation (COS) on oocyte and subsequent embryo quality remains controversial. In the present study we have compared embryo quality in natural and stimulated cycles in the same group of patients.

**METHODS:** This retrospective study was comprised of patients with a regular menstrual cycle who had IVF after COS using rFSH in a long GnRH agonist protocol. In all stimulated cycles the patients had fresh embryos transferred and surplus good quality embryos cryopreserved. Subsequently the same patients were treated with a modified FER cycle (mFER) where thawing of the frozen embryos was combined with aspiration of the dominant follicle in the natural cycle. The embryo cleavage stage and quality score were compared between the stimulated and the natural cycle for the patients having an embryo in the natural cycle.

**RESULTS:** In 177 cases patients returned for mFER in a natural cycle. Spontaneous

ovulation had occurred in 35 cycles. In 17 cycles no oocyte was retrieved at aspiration and in 125 cycles 128 oocytes were aspirated. In the stimulated cycles from these patients we had obtained 950 embryos (cleavage rate 70.4%) versus 85 embryos (cleavage rate 66.4%) ( $P = 0.34$ ) in the natural cycles. Comparing the embryos in the natural and stimulated cycles in all patients having an embryo in the natural cycle, we found no difference in the distribution between the different cleavage stages. Of the cleaved embryos, 53% in the stimulated cycles had  $>4$  cells versus 59% in the natural cycles after 2 days culture ( $P = 0.31$ ). In the stimulated cycles 61% of the embryos had  $<10\%$  fragmentation at the time of transfer on day 2, compared to 69% in the natural cycles ( $P = 0.15$ ).

**CONCLUSION:** The administration of exogenous gonadotrophins was not reflected in cleavage capacity or quality assessment of the resulting embryos.

---

**Key words:** *Developmental potential/embryo quality/impact of gonadotrophins/in vitro/in vivo*

---



## **INTRODUCTION AND LITERATURE REVIEW**

Limited information is available on the impact of gonadotrophin stimulation on the quality of oocytes and embryos. In mice, increased pre- and post-implantation mortality and fetal growth retardation have been observed in superstimulated animals (Ertzeid and Storeng, 1992). A case report by Akagbosu et al. (1998) and a paper by Aboulghar et al. (1997) suggest a negative effect of ovarian hyperstimulation syndrome on the quality of the oocytes. In a recent study Ng et al. (2003) found no negative impact on the quality of the oocytes and embryos in patients with excessive ovarian response during controlled ovarian stimulation (COS) compared to patients with a more moderate response. However, no comparison was made to embryos from natural cycles.

A number of studies have previously compared stimulated and unstimulated IVF cycles (Levy et al., 1991; Svalander et al., 1991; Paulson et al., 1992; Claman et al., 1993; MacDougall et al., 1994; Lindheim et al., 1997; Ingerslev et al., 2001; Ng et al., 2001). The studies report very different

results with implantation rates varying from 0 to 33% in the natural cycles and from 7 to 24% in the stimulated cycles. For review see Pelinck et al. (2002).

In this retrospective study we have utilized a previously described (Kim et al., 1996) treatment modality, modified frozen embryo replacement (m-FER), in which we aspirate and fertilize the oocyte from the leading follicle when monitoring the woman's natural cycle for correct timing of the FER treatment.

This means that we can compare the developmental capacity and embryo quality in a pool of embryos originating from oocytes aspirated in the same cohort of women, once at the occasion of an ovarian stimulation and once at the time of spontaneous ovulation in the same woman.

The aim of this study was to compare the morphology and early cleavage stages of the embryos from stimulated and natural cycles in the same woman in order to assess any effect of COS on early embryo development. Thus in backdrop the aim of present research paper is “**A COMPARATIVE CLINICAL EVALUATION IN NATURAL VERSUS STIMULATED IVF CYCLES EMBRYO**”



## **MATERIALS AND METHODS**

### **PATIENTS**

This retrospective analysis includes 177 couples undergoing IVF at authors word desitination Zulekha Hospital, Sharjah, UAE. The patients were referred to IVF due to tubal infertility or unexplained infertility. The study represents a consecutive series of patients returning for a modified FER cycle after a stimulated treatment cycle. The study was approved by all legal formalities.

### ***Stimulated cycles***

All patients were treated with a long protocol using GnRH agonist (Suprefact®, Hoechst Marion Roussel; or Synarela, Phamacia) as down-regulation for desensitization from day 21 of the cycle onwards and recombinant (r)FSH (Gonal-F®, Serono; or Puregon®, Organon) for COS. hCG (Profasi®, Serono) was given 36 h before oocyte retrieval. An average of 12.3  $\pm$  4.9 follicles were aspirated. While only clearly fertilized embryos (two pronuclei) were transferred, the data in this study may include a few cases with inconclusive assessment of fertilization. Embryo replacement was performed after 48 h.

### ***Modified FER cycles***

In the natural cycles we aspirated and fertilized the oocyte from the leading follicle when monitoring the woman's cycle for correct timing of the FER cycle. The aspiration was performed when the dominant follicle was >17 mm in diameter. hCG (Profasi®; Serono) was given 36 h before oocyte retrieval.

### ***Embryo culture***

The oocytes were cultured in 4-well dishes (Nunc) in 0.5 ml standard IVF medium (Universal IVF media; Medicult). After 4 h of culture, insemination was performed by addition of  $0.15 \times 10^6$  sperm cells. Embryo evaluation was performed on the morning of day 2 after insemination.

### ***Embryo evaluation***

Both in the stimulated and in the natural cycles the embryos were evaluated for cleavage stage and scored for morphology prior to transfer, in accordance with previously described criteria (Staessen et al., 1990; Ziebe *et al.*, 1997). Briefly, the scoring system was: 1.0: equally sized symmetrical blastomeres without fragmentation; 2.0: unevenly sized blastomeres without fragmentation; 2.1: embryos with <10% fragmentation; 2.2: embryos with 10-20% fragmentation; 3.0:



embryos with 20-50% fragmentation; 4.0: embryos with >50% fragmentation.

### STATISTICAL ANALYSIS

Statistical analysis was done by the  $\chi^2$ -test. Values was considered significant if  $P < 0.05$ .

### RESULTS

The study included a total of 177 stimulated cycles and 177 subsequent natural cycles (Table IA). In 35 natural cycles ovulation had taken place prior to aspiration. In another 17 natural cycles no oocytes were aspirated. In 125 natural cycles oocytes were aspirated (Table IA).

In the stimulated cycles from women with oocytes in the natural cycle, a

total of 1349 oocytes were aspirated ( $10.8 \pm 4.7$  oocyte/cycle). In the natural cycles 128 oocytes were aspirated in 125 cycles (Table IB). The average age of these women was  $32.8 \pm 3.2$  at the time of the stimulated cycle and  $33.6 \pm 3.2$  when they returned for the m-FER cycle.

No significant difference was found in cleavage rate between oocytes from stimulated and natural cycles. In the stimulated cycles we obtained 950 embryos (cleavage rate 70.4%), versus 85 embryos (cleavage rate 66.4%) in the natural cycles ( $P = 0.34$ ) (Table IB).

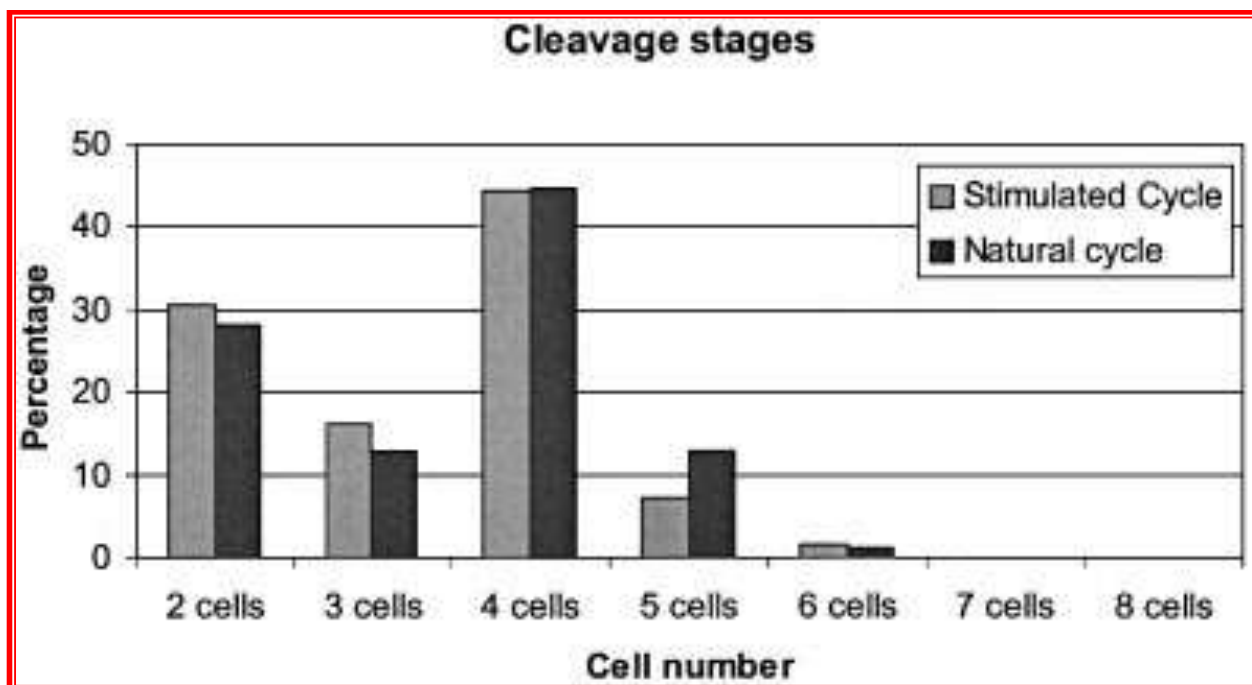
Table I.

|  | Stimulated cycles | Natural cycles |
|--|-------------------|----------------|
| <b>Table I A. All patients</b>                               |                   |                |
| <b>No. of included patients</b>                              | 177               | 177            |
| <b>Mean age of the women (years)</b>                         | $32.8 \pm 3.3$    | $33.6 \pm 3.3$ |
| <b>Average total FSH dose in the stimulated cycle (IU)</b>   | $1.881 \pm 807$   | -              |
| <b>Average days of stimulation</b>                           | $10.4 \pm 2.6$    | -              |
| <b>Average endometrial thickness (mm)</b>                    | $11.0 \pm 7.8$    | $8.2 \pm 1.9$  |
| <b>Day of hCG</b>  | $11.4 \pm 1.9$    | $13.2 \pm 2.2$ |
| <b>No. of cycles</b>   | 177               | 177            |
| <b>Ovulation prior to aspiration</b>                         | 0                 | 35             |
| <b>No. of cycles with no oocyte retrieved</b>                | 0                 | 17             |
| <b>No. of cycles with oocytes</b>                            | 177 (100)         | 125 (71)       |
| <b>Table I B. Patients with oocytes in the natural cycle</b> |                   |                |



|  |            |                          |
|--|------------|--------------------------|
| No. of cycles with oocytes                               | 125        | 125                      |
| Mean age of woman at treatment (years)                   | 32.8 ± 3.2 | 33. ± 6 3.2              |
| No. of oocytes retrieved                                 | 1349       | 128                      |
| No. of embryos   | 950        | 85                       |
| Cleavage rate (%)  | 70.4       | 66.4, <i>P</i> = 0.34    |
| <b>Table I C. Patients with embryos in natural cycle</b> |            |                          |
| No. of cycles with embryos                               | 85         | 85                       |
| Mean age of woman at treatment (years)                   | 32.8 ± 3.4 | 33.7 ± 3.3               |
| No. of oocytes   | 920        | 85                       |
| No. of embryos   | 670        | 85                       |
| No. of embryos with >4 cells                             | 355 (53)   | 50 (59), <i>P</i> = 0.31 |
| No. of embryos with <10% fragmentation                   | 411 (61)   | 59 (69), <i>P</i> = 0.15 |

*Values in parentheses are percentages.*



**Figure 1.** Cleavage stage of all embryos after 2 day culture of oocytes from stimulated and natural cycles in patients having an embryo in the natural cycle.

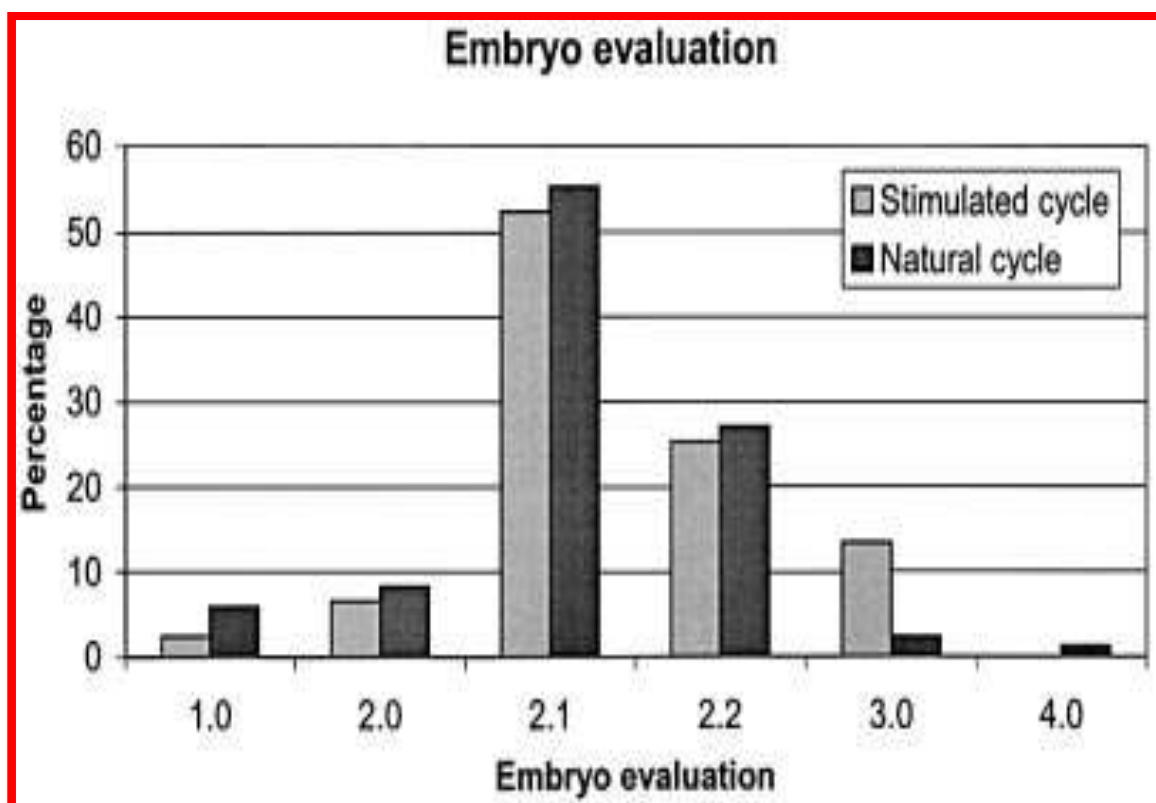
In order to perform a relevant comparison of embryo cleavage stage and quality score

between the stimulated and natural cycles, we have compared all patients having an



embryo in the natural cycle. The mean age of the women in this subgroup was  $32.8 \pm 3.4$  years in the stimulated cycle and  $33.7 \pm 3.3$  years in the natural cycle (Table IC). The distribution between the different cleavage stages was the same in both groups (Figure 1). After 2 days of culture

44.3% of the embryos had cleaved to the 4-cell stage in the stimulated cycles versus 44.7% in the natural cycles. Furthermore, 53% of the embryos in the stimulated cycles had >4 cells at day 2 compared to 59% in the natural cycles ( $P = 0.31$ ) (Table IC).



*Figure 2. Evaluation of all embryos after 2 day culture of oocytes from stimulated and natural cycles in patients having an embryo in the natural cycle.*

We found no difference in the fragmentation pattern between the stimulated and natural cycles (Figure 2). In the stimulated cycles, a total of 61% of the embryos had <10% fragmentation at the

time of transfer, compared to 69% in the natural cycles ( $P = 0.15$ ) (Table IC).

## DISCUSSION

The data from the present retrospective analysis suggest that the



administration of recombinant FSH after pituitary desensitization is not reflected in the morphology of the early stages of embryonic development. Neither the cleavage rate of the oocytes nor the early cleavage stage morphology assessment of the embryos differed between the stimulated cycles and the natural cycles.

These findings suggest that the only reason why hormonal stimulation increases the likelihood of achieving a pregnancy is by increasing the number of oocytes retrieved which again might provide us with more embryos to select from at the time of transfer.

In theory, the hormonal environment in the natural cycle provides optimal conditions for the maturing follicle and oocyte. Indeed cycles with desensitization followed by stimulation with gonadotrophins differs markedly from the natural cycle. In a study by Kaneko et al. (2000), COS was shown to have an impact on apoptosis in the granulosa cells.

During IVF we are often faced with pronounced embryo heterogeneity both in terms of morphology (Cummins *et al.*, 1986; Puissant *et al.*, 1987; Claman *et al.*, 1987; Staessen *et al.*, 1992; Steer *et*

*al.*, 1992; Shulman *et al.*, 1993; Giorgetti *et al.*, 1995; Ziebe *et al.*, 1997; Hardarson *et al.*, 2001; Van Royen *et al.*, 2001) and chromosomal constitution (Ziebe *et al.*, 2003; Johansson *et al.*, 2003). The reason for this diversity is largely unknown but it reflects to some degree the developmental capacity of the embryo. Several underlying factors are probably contributing to the diversity including the in vitro environment and culture conditions. In a previous study Van Blerkom and Davis (2001) demonstrated the effect of repeated ovarian stimulation in mice, resulting in a significant increase in the frequency of spindle defects resulting in chromosomal errors with each series of ovarian stimulation.

*We have no directly measurable parameters of oocyte quality except for the nuclear maturity. However, an indirect indication of the oocyte quality could be the developmental capacity.*

*Despite the fact that no difference was found in this study concerning the light microscopic morphological appearance of the embryos, there may still be an effect as a result of administration of gonadotrophins on embryo quality not reflected until*



*implantation or even later. The advantage of our study design was that the stimulated and the natural cycles were in the same patients.* However, the study design with stimulated versus natural cycles resulting in many versus one embryo in each woman this retrospective study may result in insufficient power to detect a minor impact on embryo development.

A disadvantage of the study design is that we were unable to assess the implantation rate of the natural cycle embryos separately, since in the majority

of cases they were transferred in combination with the thawed embryos.

*In conclusion, we find that the use of hormonal stimulation in assisted reproductive treatment does not alter the ability of the oocyte to cleave after fertilization.* Furthermore, the developmental capacity, defined as the number of blastomeres after 2 days of culture, is unaffected of the use of exogenous gonadotrophins. Finally, embryo quality in terms of degree of fragmentation is similar between embryos resulting from stimulated cycles and natural cycles.

## REFERENCES

- Aboulghar MA, Mansour RT Serour GI, Ramzy AM and Amin YM (1997) Oocytes quality in patients with severe ovarian hyperstimulation syndrome. *Fertil Steril* 68,1017±1021.
- Akagbosu A, Marcus S, Abusheikha N, Avery S and Brinsden P (1998) Does ovarian hyperstimulation syndrome affect the quality of oocytes. *Hum Reprod* 13,2583±2584.
- Claman P, Armant DR, Seibel MM, Wang T-A, Oskowitz SP and Taymor ML (1987) The impact of preembryo quality and quantity on implantation and the establishment of viable pregnancies. *J In Vitro Fertil Preembryo Transfer* 4,218±222.
- Claman P, Domingo M, Garner P et al (1993) Natural cycle in vitro fertilization-embryo transfer at the University of Ottawa: an inefficient therapy for tubal infertility. *Fertil Steril* 60,298±302.
- Cummins JM, Breen TM, Harison KL, Shaw JM, Wilson LM and Hennessey JF (1986) A formula for scoring human preembryo growth rates in in vitro fertilization:





its value in predicting pregnancy and in comparison with visual estimates of preembryo quality. *J In Vitro Fertil Preembryo Transfer* 3,284±295.

- Ertzeid G and Storeng R (1992) The impact of ovarian stimulation on implantation and fetal development in mice. *Hum Reprod* 16,221±225.
- Giorgetti C, Terriou P, Auquier P, Hans E, Spach J-L, Salzmänn J and Roulier R (1995) Preembryo score to predict implantation after in-vitro fertilization: based on 957 single preembryo transfers. *Hum Reprod* 10,2427±2431.
- Hardarson T, Hanson C, Sjöëgren A and Lundin K (2001) Human preembryos with unevenly sized blastomeres have lower pregnancy and implantation rates: indication for aneuploidy and multinucleation. *Hum Reprod* 16,313± 318.
- Ingerslev HJ, Højgaard A, Hindkjær J and Kesmodel U (2001) A randomized study comparing IVF in the unstimulated cycle with IVF following clomiphene citrate. *Hum Reprod* 16,696±702.
- Johansson M, Hardarson T and Lundin K (2003) There is a cutoff limit in diameter between a blastomere and a small anucleate fragment. *J Assist Reprod Genet* 20,309±313.
- Kaneko T, Saito H, Takahashi T, Ohta N, Saito T and Hiroi M (2000) Effect of controlled ovarian hyperstimulation on oocyte quality in terms of the incidence of apoptotic granulosa cells. *J Assist Reprod Genet* 17,580±585.
- Kim SH, Kim CH, Suh CS, Moon SY, Lee JY and Chang YS (1996) Simultaneous program of natural-cycle in vitro fertilization and cryopreserved-thawed embryo transfer. *J Assist Reprod Genet* 13,716±721.
- Levy MJ, Gindoff P, Hall J and Stillman RJ (1991) The efficacy of natural versus stimulated cycle IVF-ET. *Fertil Steril* 56(Suppl),S15±16.
- Lindheim SR, Vidali A, Ditkoff E and Sauer MV (1997) Poor responders to ovarian hyperstimulation may benefit from an attempt at natural-cycle oocyte retrieval. *J Assist Reprod Genet* 14,174±176.



- MacDougall MJ, Tan SL, Hall V et al (1994) Comparison of natural with clomiphene citrate-stimulated cycles in in vitro fertilization: a prospective, randomized trial. *Fertil Steril* 61,1052±1057.
- Ng EHY, Chui DKC, Tang OS et al (2001) In vitro fertilization and embryo transfer during natural cycles. *J Reprod Med* 46,95±99.
- Ng EHY, Lau EYL, Yeung WSB and Ho PC (2003) Oocyte and embryo quality in patients with excessive ovarian response during in vitro fertilization treatment. *J Assist Reprod Genet* 20,186±191.
- Paulson RJ, Sauer MV, Francis MM et al (1992) In vitro fertilization in unstimulated cycles: the University of Southern California experience. *Fertil Steril* 57,290±293.
- Pelinck MJ, Hoek A, Simons AHM and Heineman MJ (2002) Efficacy of natural cycle IVF: a review of the literature. *Hum Reprod Update* 2,129± 139.
- Puissant F, Van Rysselberge M, Barlow P, Deweze J and Leroy F (1987) Preembryo scoring as a prognostic tool in IVF treatment. *Hum Reprod* 2,705±708.
- Shulman A, Ben-Nun I, Ghetler Y, Kaneti H, Shilon M and Beyth Y (1993) Relationship between preembryo morphology and implantation rate after in vitro fertilization treatment in conception cycles. *Fertil Steril* 60,123±126.
- Staessen C, Van den Abbeel E, Carle M, Khan I, Devroey P and Van Steirteghem AC (1990) Comparison between human serum and Albuminar-200 supplement for in-vitro fertilization. *Hum Reprod* 5,336± 341.
- Staessen C, Camus M, Bollen N, Devroey P and Van Steirteghem AC (1992) The relationship between preembryo quality and the occurrence of multiple pregnancies. *Fertil Steril* 57,626±630.
- Steer CV, Mills CL, Tan SL, Campbell S and Edwards RG (1992) The cumulative embryo score: a predictive embryo scoring technique to select the optimal number of embryos to transfer in an in-vitro fertilization and embryo transfer programme. *Hum Reprod* 7,117±119.
- Svalander P, Green B, Haglund K et al (1991) Natural versus stimulated cycles in IVF-ET treatment for tubal factor infertility. *Hum Reprod* 6(Suppl 1),101±102.



- VanBlerkom J and Davis P (2001) Differential effects of repeated ovarian stimulation on cytoplasmic and spindle organization in metaphase II mouse oocytes matured in vivo and in vitro. Hum Reprod 16,757±764.
- VanRoyen E, Mangelschots K, De Neubourg D, Laureys I, Ryckaert G and Gerris J (2001) Calculating the implantation potential of day 3 embryos in women younger than 38 years of age: a new model. Hum Reprod 16,326± 332.
- Ziebe S, Petersen K, Lindenberg L, Andersen A-G, Gabrielsen A and Nyboe Andersen A (1997) Embryo morphology or cleavage stages: how to select the best embryos for transfer after in vitro fertilization. Hum Reprod 12,1545±1549.
- Ziebe S, Bech B, Petersen K, Mikkelsen AL, Gabrielsen A and Nyboe Andersen A (1998) Resumption of mitosis during post-thaw culture: a key parameter in selecting the right embryos for transfer. Hum Reprod 13,178±181.
- Ziebe S, Lundin K, Loft A, Bergh C, Nyboe Andersen A, Selleskog U, Gründahl C and Arce J-A (2003) Embryo morphology and total chromosomal constitution for chromosome 3, 16, 18, 21, 22, X and Y after 3 days of culture of 144 randomly selected donated human oocytes. Hum Reprod 18,2575±2581.