GLAUCOMA AND NEUROPROTECTION:

THERAPEUTIC PERSPECTIVES

ITALO GIUFFRE' MD PhD

Department of Ophthalmology (Head: Prof. A. Caporossi) – Catholic University of Rome – Rome – ITALY – EU.

ABSTRACT

Glaucoma is the main cause of irreversible blindness worldwide. It affects approximately 70 millions of people in the world. It is an optic neuropathy. It is characterized by a functional damage, caused by progressive retinal ganglion cells (RGC) and their axons progressive loss (1). It is caused by a high intraocular pressure (IOP) which is responsible of RGC death in open-angle glaucoma. There is also a low-tension glaucoma. The second risk factor of this disease is the age. It is higher every decade over 40 years old (2-4). Other risk factors are: Afro-american race, myopia, diabetes, positive family anamnesis. It is actually unknown why an increase of IOP may cause a thinning of retinal nervous fiber layer (RNFL), a cupping of optic disc and typical visual field damage. There are two aetiopathogenetic theories: mechanical and vascular ones (5,6). Modern research is exploring which molecular mechanisms upgrade or downgrade both mechanical and vascular damages even when IOP is in a normal range (7,8). We hope to stop the neural cell damage by using neuroprotective agents associated to conventional hypotonizing drugs.

KEY-WORDS: glaucoma, neuroprotection, optic disc, retinal nerve fiber layer.

The Author declares that no competing interests exist.

GLAUCOMA AND NEUROPROTECTION: THERAPEUTIC PERSPECTIVES

In glaucoma disease there is apoptosis either in animal model or in neuron cells (9,10) or in normal-tension glaucoma patients (11,12). It is not yet clear how the increase of IOP may trigger the apoptosis. Gregory M.S. et al. (13) proved the role of Fas Ligand (FasL). It is a transmembrane type 2 protein, belonging to TNF family. It is present in the ocular tissues and it contributes to the privileged immunological state. It prevents the angiogenesis and it triggers the apoptosis of the inflammatory cells (14). FasL is involved in the RGC T-cell mediated death (15,16). FasL activities depend if it binds to the cell membrane or if it is soluble. There are two FasL forms: the T-released microvescicles (mFasL) longer than a soluble form truncated by metalloproteinases in the cell membrane (17,18,19). In their experiment Gregory M.S. et al. (13) showed that when FasL is present, the RGC death is faster. Although, when FasL is not present, the RGC death stopped. Metalloproteinases (MP3 and MP7) truncate FasL (20). The IOP increase may increase the level of α -TNF, responsible for the anomaly of MP and their tissue inhibitors (TIMPs). In a previous study RGCs and glial cells were studied. The glial cells produce α-TNF and NO, responsible for the apoptosis of the ganglion cells. It happened after ischemia and increase of hydrostatic pressure. Apoptosis was reduced by 66% by a neutralizing α -TNF antibody and by 50% of an inducible nitric oxide synthetasis (iNOS) (21).

These results show that the glial cells are neurotoxic after ischemia or increased hydrostatic pressure. α-TNF is a neurotoxic agent on the ganglion cells. Their antagonist may be neuroprotective, considering that α -TNF may produce NO (22,23,24). In vivo an increase of NO is present in open-angle glaucoma, normal-tension glaucoma and pseudoexfoliative glaucoma patients (25,26). Cytokine is longlasting and it is responsible of the neuronal loss even when the IOP is normal (27).

Another protein, beyond α -TNF, is produced by neural microglial cells when the IOP is high. It causes RGC death and it is called α2macroglobulin (α2M). Its gene is upregulated after 7 days of increased IOP and it lasts 20 days and it is responsible of RGC death (28,29,30). Bay et al. (31) proved the presence of α 2M in the aqueous humor of either experimentally induced or in glaucoma patients. This protein is produced in the retina but it is present in the anterior chamber and it is a marker of ocular hypertension. α2M seems to neutralize the Nerve Growth Factor (NGF), protective neurotrophin of RGC upregulating TrkA receptors. This data explains why only high doses of NGF protect RGCs (32,33,34).

Guo et al. (35) supported the hypothesis that the increase of IOP induces the apoptosis of RGC, remodelling the extracellular matrix (ECM) and the optic nerve by lamina cribrosa axonal compression, axonal flux stop and then death. In vivo they showed a positive correlation between the increase of IOP and upregulation of MMP9 (p<0.001), TIMp-1 (p<0.05) and TGFβ-2 (p<0.05). MMP9 is positive correlated to the RGC apoptosis ($p<0.001$) and laminin loss ($p<0.01$). On the optic nerve head type 1 collagen increased $(p<0.01)$.

These evidences are supported by an increase of MMP-9 associated to apoptosis in CNS (36) and a reduction of laminin (37,38).

The increase of MPP-9 may be the consequence of the mechanical effect of IOP on retinal ganglion axons and an increase of neurotransmitter glutammate after increased IOP, ischemia and damage (39,40,41,42).

A Monthly Double-Blind Peer Reviewed Refereed Open Access International e-Journal - Included in the International Serial Directories **International Journal of Research in Medical and Basic Sciences (IJRMS)** http://www.mbsresearch.com email id- irjmss@gmail.com Page 63

The increase of TIMP-1 counteractes MMP9 (43). That's why it is important to stop the apoptotic cascade in the therapy of glaucoma. Autophagy is important in neuronal cell death in glaucoma.

In the eukariotes, autophagy is a physiological process of degradation of proteins, cytoplasmic organelles and in lysosomes. It may induce tissue remodelling after ischemia and retinal neurodegenerative diseases (44,45,46).

Piras et al. studied autophagy in ganglion cell neurons after ischemia/reperfusion in murine retinas. Autophagic markers, LC3 and LAMP1 increased after 12 and 24 hours. It was strongly stopped by 3-metiladenine, a strong inhibitor of autophagosomes (47).

Ischemia is an important clinical problem, shared by glaucoma and both diabetic and hypertensive retinopathy.

Fluctuation of retinal extracellular pH was associated to ASICs (Acid-sensing ion channels). ASIC1a is a Ca^{2+} and Na⁺ channel (48,49). In culture, ASIC1a upregulated after oxygen deprivation and an increase of intracellular Ca^{2+} . Amiloride and psalmotoxin 1 reduces RGCs death in vivo (50). This may be a target of neuroprotection in glaucomatous, diabetic and hypertensive patients.

Research focused on mithocondria, oxidative stress (51) and inflammation which cause the disease by neuronal death.

Liu Q. (2011, 52) evaluated 4-idrossi-2-nonenale (HNE) and heme-oxygenasis 1 (HO-1) in a retinal ganglion cell culture after 0, 30, 60 or 100 mmHg hydrostatic pressure for 2 hours and a murine model after increase of IOP at 30, 60 and 100 mmHg as long as 1 hour. Both experiments increased HNE and HO-1. HNE induced neuronal death .

Even photoreceptors may die during glaucoma but RGC are more sensitive (53).

An antioxidant, such as resveratrol, inhibits the oxidative stress and it may be useful in the neuroprotection of glaucoma.

Flavonoids showed a neuroprotective effect against oxidative stress, glutammate and hypoxia (54).

These drugs have a neuroprotective effect in cancer, cardiovascular and neurodegenerative diseases (55,56,57,58,59,60,61).

Nicotriflorin, rutin and quercitin have a neuroprotective effect. They are present in fruit and vegetables. Quercitin is present in tea. In a media, flavonoids were tested by inhibiting caspasis 3 and calpain (a cystein-proteinasis). Nicotriflorin and rutin were neuroprotective against hypoxia, glutammate and oxydative stress at a concentration of 1nM, while quercitin needed higher concentration (100 nM).

The flavonoid neuroprotection depends on their nucleus. They influence the glutathione metabolism, antioxidant facility and keep low Ca^{2+} concentration notwithstanding high free oxygen radicals (FOR) levels (62,63).

A Monthly Double-Blind Peer Reviewed Refereed Open Access International e-Journal - Included in the International Serial Directories **International Journal of Research in Medical and Basic Sciences (IJRMS)** http://www.mbsresearch.com email id- irjmss@gmail.com Page 64

The flavonoids are useful as neuroprotectant in the oxydative stress by hypoxia and glutammate damage may depend on the influence of hypoxia and glutammate in the cell death.

Another factor involved in ganglion cell damage during glaucoma, diabetic retinopathy and ischemia is the light.

Li G. Y. et al. (64) proved that light may cause retinal ganglion cell damage destroying DNA and mitochondria.

RGC-5 cells were used (65). 2600 lux light may damage DNA and activate poly-ADP-ribose polymerase 1 (PARP-1). It acts to repair small DNA damage. Instead, PARP-1 inhibition is neuroprotective against cell damage. Benzamide and nicotinamide were used as PARP-1 inhibitor, increasing the cell survival from 25.4 to 38.1% at a concentration of 10 mM. NU1025, a new inhibitor, increased the cell survival by 77.1%. Also Nfenilmaleimide, an inhibitor of AIF (apoptosis inducing factor) increased the cells from 15.72 to 29.35%. It is a proapoptotic factor, anchored in the inner mithocondrial membrane. It binds to DNA and RNA and it causes a damage to caspasis-indipendent chromatine and DNA fragmentation.

Light increases ten times the Ca^{2+} intracellular concentration and cobaltus, an inhibitor of this ion, reduces the cell death, caused by inhibition of ATP production and NO production (66,67).

Mitochondria are involved in the ganglion cell death in glaucoma. The light may act on cycloxygenasis, P450 cytochrome and flavin, producing the perossinitrite anion (ONOO, 68).

Osborne N.N. (69), in order to neuroprotect against glaucoma, advices some drugs as creatine, αlipoic acid, nicotinammide and epigallocatechingallate (ECGC).

Wang Y.S. et al. (70) recently studied the effect of Ginkgo biloba (EGb761) on the ganglion cell culture, associated to glutammate. In the control the survival was $61.94\pm7.75\%$, in glutammate culture 44.59±4.19%, in EGb761 75.05±3.90% and 63.19±9.44% in both glutammate and EGb761 culture.

Neuroglobin (NGB) is an endogenous neuroprotector. In an experimental murine glaucoma, it was induced by high IOP level to protect the RGCs. It reduced the superoxyde and ATP production. This globin may be a therapeutic target against glaucoma.

In 2011 more and more researches tried to identify new neuroprotective factors. Q.L. (71) found a new soluble antagonist NOGO-66 (sNgR-Fc), to stop the RGCs death at 5 days, 2 and 4 weeks (72).

Optineurin gene is expressed in RGC. Its mutation may be associated to open-angle glaucoma and amyotrophic lateral sclerosis. Neurotrophin 3 (NT-3) is a neuroprotective antagonist (73).

P53 and cycline-dependent kinase 5 (CDK-5), responsible of neurodegenerative diseases, increases p-N32A (S1232), one of N-methyl-D-aspartate (NMDA) receptors (74). In glaucoma, its activation causes RGC apoptosis (75).

IOP-independent neuroprotection was identified also in prostaglandin analogues such as latanoprost, tafluprost and bimatoprost, at a concentration of 100 nM (76,77).

A Monthly Double-Blind Peer Reviewed Refereed Open Access International e-Journal - Included in the International Serial Directories **International Journal of Research in Medical and Basic Sciences (IJRMS)** http://www.mbsresearch.com email id- irjmss@gmail.com Page 65

Apart from neuroprotection, glaucoma is surely associated to ageing. Scanning electronic microscope (SEM) was very useful in identifying differences associated to ageing (p<0.001) in young and adult retinas about retinal thickness, ganglion cells, capillaries and synaptic junctions numbers (78).

Chierzi S. et al. (79) showed that the ability of cone regeneration depends on the ageing and inhibition of ERK (extracellular-signal regulated kinase 1,2) and Protein-kinase A (PKA).

These concepts may be useful in the neuroprotective therapy of glaucoma, optic neuropathy and other retinopathies. Further studies are needed to find oral pharmacological associations useful to neuroprotect the ganglion cells, bioavailable and without side-effects.

REFERENCES

- **1.** Weinreb R.N., Khaw P.T. Primary open-angle glaucoma. Lancet 2004; 363: 1711-20.
- **2.** Kwon Y.J, Fingert J.H., Kuhen M.H. et al.. Primary open-angle glaucoma. N. Engl. J. Med. 2009, 360: 1113-24.
- **3.** Armaly M.F., Krueger D.E., Maunder L. et al.. Biostatistical analysis of the collaborative glaucoma study. 1. Summary report of the risk factors for glaucomatous visual field defects. Arch. Ophthalmol. 1980; 98: 2163-71.
- **4.** Guedes G., Tsai J.C., Loewen N. A. Glaucoma and aging. Curr. Aging Sci. 2011; 4: 110-7.
- **5.** Leske M.C. The epidemiology of open-angle glaucoma: a review. Am. J. Epidemiol. 1983;118: 166-91.
- **6.** Halpern D.L., Grosskreutz C.L.. Glaucomatous optic neuropathy: mechanisms of disease. Ophthalmol. Clin. North Am. 2002; 15: 61-8.
- **7.** Yoles E., Schwartz M. Potential neuroprotective therapy for glaucomatous optic neuropathy. Surv. Ophthalmol. 1998; 42: 367-72.
- **8.** Rudzinski M., Saragovi H.U. Glaucoma: validated and facile in vivo experimental models of a chronic neurodegenerative disease for drug development. Curr. Med. Chem. 2005; 5: 43-9.
- **9.** Agar A., Li S., Agarwal N. et al.. Retinal ganglion cells line apoptosis induced by hydrostatic pressure. Brain Res. 2006; 1086: 191-200.
- **10.** Agar A., Yip S.S., Hill M.A. et al. Pressure related apoptosis in neuronal cell lines. J. Neurosci. Res. 2000; 60: 495-503.
- **11.** Garcia-Valenzuela E., Shareef S., Walsh J. et al.. Programmed cell death of retinal ganglion cells during experimental glaucoma. Exp. Eye Res. 1995; 61: 33-44.
- **12.** Tatton N.A., Tezel G. Insolia S.A. et al. In situ detection of apoptosis in normal pressare glaucoma: a preliminary examination. Surv. Ophthalmol. 2001; 45: S268-S272.
- **13.** Gregory M.S., Hackett C.G., Abernathy E.F. et al.. Opposing roles for membrane bound and soluble Fas-ligand in glaucoma-associated retinal ganglion cell death. PLoS ONE 2011; 6: e17659.
- **14.** Lee H.O., Ferguson T.A.. Biology of FasL. Cytokine Growth Factor Rev. 2003; 14: 325-35.
- **15.** Wax M.B., Tezel G., Yang J. et al.. Induced autoimmunity to heat shock proteins elicits glaucomatous loss of retinal ganglion cell neurons via activated T-cell-derived fas-ligand. J. Neurosci. 2008; 28: 12085-96.

A Monthly Double-Blind Peer Reviewed Refereed Open Access International e-Journal - Included in the International Serial Directories **International Journal of Research in Medical and Basic Sciences (IJRMS)** http://www.mbsresearch.com email id- irjmss@gmail.com Page 67

- **16.** Ju K.R., Kim H.S., Kim J.S. et al.. Retinal glial cells responses and Fas/FasL in rats with chronic ocular hypertension. Brain Res. 2006; 1122: 209-21.
- **17.** Tanaka M., Suda T., Takahashi T. et al.. Expression of the solubile form of the human Fas ligand in activated lymphocytes. EMBO J. 1995; 14: 1129-35.
- **18.** Jodo S., Xiao S., Hohlbaum A.M. et al.. Apoptosis-inducing membrane vesicles: a novel agent with unique properties. J. Biol. Chem. 2001; 276: 39938-44.
- **19.** Bossi G., Griffiths G.M.. CTL secretory lysosomes: biogenesis and secretion of a harmful orfanelle. Semin. Immunol. 2005; 17: 87-94.
- **20.** Vargo-Gogola T., Crawford H.C., Fingleton B. et al.. Identification of novel matrixmetalloproteinase-7 (matrylisin) cleavage sites in murine and human Fas-ligand. Arch. Biochem. Biophys. 2002; 408: 155-61.
- **21.** Tezel G., Wax M.B.. Increase production of necrosis tumor factor-α by glial cells exposed to simulated ischemia or elevated hydrostatic pressure induces apoptosis in cocultured retinal ganglion cells. The Journal of Neuroscience 2000; 20: 8693-700.
- **22.** Goureau O., Amiot F., Dautry F. et al.. Control of nitric oxide production by endogenous TNF- $α$ in mouse retinal pigmented epithelial and Muller glial cells. Biochem. Biophys. Res. Commun. 1997; 240: 132-5.
- **23.** Teneka M.T., Loschmann P.A., Gleichmann M. et al.. Induction of nitric-oxide-mediated apoptosis in neuronal PC12 cells after stimulation with tumor necrosis factorα/lipopolysaccharide. J. Neurochem. 1998; 71: 88-94.
- **24.** Shafer R.A., Murphy S.. Activated astrocytes induce nitric oxide synthase-2 in cerebral endothelium via tumor necrosis factor-α. Glia 1997; 21: 370-9.
- **25.** Sawada H., Fukuchi T., Tanaka T. et al.. Tumor necrosis factor-α concentrations in the aqueous humor of patients with glaucoma. Invest. Ophthalmol. Vis. Sci. 2010; 51: 903-6.
- **26.** Tezel G., Li L.Y, Patil R.V. et al.. TNF-α and TNF-α receptor-1 in the retina of normal and glaucomatous eyes. Invest. Ophthalmol. Vis. Sci. 2001, 42: 1787-94.
- **27.** McKinnon S.J.. Glaucoma: ocular Alzheimer's disease? Front Biosci. 2003; 8: S1140-56.
- **28.** Shi Z., Rudzinski M., Meerovitch K. et al. Alpha-2 macroglobulin is a mediator of retinal ganglion cell death in glaucoma. J. Biol. Chem. 2008; 283: 29156-65.
- **29.** Bay Y., Dergham P., Nedev H. et al. In chronic and in acute models of retinal neurodegeneration TrkA activity is neuroprotective while P75NTR activity is neurotoxic through a paracrine mechanism. J. Biol. Chem. 2010; 285: 39392-400.

A Monthly Double-Blind Peer Reviewed Refereed Open Access International e-Journal - Included in the International Serial Directories **International Journal of Research in Medical and Basic Sciences (IJRMS)** http://www.mbsresearch.com email id- irjmss@gmail.com Page 68

- **30.** Bay Y., Shi Z., Zhuo Y. et al.. In glaucoma the up-regulated truncated TrkC. T1 receptor isoform in glia causes increased TNF-alpha production, leading to retinal ganglion cell death. Invest. Ophthalmol. Vis. Sci. 2010; 51: 6639-51.
- **31.** Bay Y., Sivori D. Woo S.B. During glaucoma, α2-Macroglobulin accumulates in aqueous humor and binds to nerve growth factor, neutralizing neuroprotection. Invest. Ophthalmol. Vis. Sci. 2011; 52: 5260-5.
- **32.** Shi Z., Birman E., Saragovi H.U.. Neurotrophic rationale in glaucoma: a TRK-A agonist, but not NGF or a p75 antagonist, protects retinal ganglion cells in vivo. Dev. Neurobiol. 2007; 67: 884- 94.
- **33.** Saragovi H.U., Hamel E., Di Polo A.. A neurotrophic rationale for the therapy of neurodegenerative disorders. Current Alzheimer research 2009; 6: 419-23.
- **34.** Lambiase A., Aloe L., Centofanti M. et al.. Experimental and clinical evidence of neuroprotection by nerve growth factor eye drops: implications for glaucoma. Proc. Natl. Acad. Sci. U.S.A. 2009; 106: 13469-74.
- **35.** Guo L., Moss S.E., Alexander R.A. et al.. Tetinal ganglion cell apoptosis in glaucoma is related to intraocular pressure and IOP-induced effects on extracellular matrix. Invest. Ophthalmol. Vis. Sci. 2005; 46: 175-82.
- **36.** Vaillant C., Meissirel C., Mutin M. et al. MMP-9 deficiency affects axonal outgrowth, migration and apoptosis in the developing cerebellum. Mol. Cell Neurosci. 2003; 24: 395-408.
- **37.** Grossmann J..Molecular mechanism of "detachment-induced apoptosis-anoikis". Apoptosis 2002; 7: 247-60.
- **38.** Tsirga S.E., Rogove A.D., Strickland S.. Neuronal cell death and tPA. Nature 1996; 384: 123-4.
- **39.** Arundine M., Tymianski M.. Molecular mechanism of glutamate-dependent neurodegeneration in ischemia and traumatic brain injury. Cell Mol. Life Sci. 2004; 61: 657-68.
- **40.** Martin K.R., Levkovitch-Verbin H., Valenta D. et al.. Retinal glutamate transporter changes in experimental glaucoma and after optic nerve transection in the rat. Invest. Ophthalmol. Vis. Sci. 2002; 43: 2236-43.
- **41.** Vorwerk C.K., Naskar R., Schuettauf F. et al.. Depression of retinal glutamate transporter function leads to elevated intravitreal glutamate levels and ganglion cell death. Invest. Ophthalmol. Vis. Sci. 2000; 41: 3615-21.
- **42.** Zhang X., Cheng M., Chintala S.K.. Kainic acid-mediated upregulation of matrix metalloproteinase-9 promotes retinal degeneration. Invest. Ophthalmol. Vis. Sci. 2004; 45: 2374- 83.

A Monthly Double-Blind Peer Reviewed Refereed Open Access International e-Journal - Included in the International Serial Directories **International Journal of Research in Medical and Basic Sciences (IJRMS)** http://www.mbsresearch.com email id- irjmss@gmail.com Page 69

- **43.** Tan H.K., Heywood D., Ralph G.S. et al.. Tissue inhibitor of metalloproteinase 1 inhibits excitotoxic cell death in neurons. Mol. Cell Neurosci. 2003; 22: 98-106.
- **44.** Levine B., Klionsky D.J.. Development by self-digestion: molecular mechanisms and biological functions of autophagy. Dev. Cell 2004; 6: 464-73.
- **45.** Qu X., Zou Z., Sun Q. et al. Autophagy genedependent clearance of apoptotic cells during embryonic development. Cell 2007; 128: 931-46.
- **46.** Adhami F., Liao G., Morozov Y.M. et al.. Cerebral ischemia-hypoxia induces intravascular coagulation and autophagy. Am. J. Pathol. 2006; 169: 566-83.
- **47.** Piras A., Gianetto D., Conte D..et al. Activation of autophagy in a rat model of retinal ischemia following high intraocular pressure. PLoS ONE 2011; 6: e22514.
- **48.** Waldmann R., Bassilana F., De Weille J. et al.. Molecular cloning of a noninactivating protongated Na+channel specific for sensory neurons. J. Biol. Chem. 1997; 272: 20975-8.
- **49.** Yermolaieva O., Leonard A.S., Schnizler M.K. et al.. Extracellular acidosis increases neuronal cell calcium by activating acid-sensing ion channel 1a. Proc. Natl. Acad. Sci. U.S.A. 2004; 101: 6752-7.
- **50.** Tan J., Ye X., Xu Y. et al.. Acid-sensing ion channel 1a is involved in retinal ganglion cell death induced by hypoxia. Mol. Vision 2011; 17: 3300-8.
- **51.** Moreno M.C., Campanelli J., Sande P. et al.. Retinal oxidative stress induced by high intraocular pressure. Free Radic Biol. Med. 2004; 37: 803-12.
- **52.** Liu Q., Ju W.K., Crowston J.G. et al.. Oxidative stress is an early event in hydrostatic pressureinduced retinal ganglion cell damage. Invest. Ophthalmol. Vis. Sci. 2007; 48: 4580-9.
- **53.** Weinreb N.R., Khaw P.T.. Primary open-angle glaucoma. Lancet 2004; 363: 1711-20.
- **54.** Nakayama M., Aihara M., Chen Y.N.. Neuroprotective effects of flavonoids on hypoxia-, glutamate- and oxidative stress-induced retinal ganglion cell death. Molecular Vision 2011; 17: 1784-93.
- **55.** Ross J.A., Kasum C.M.. Dietary flavonoids: bioavailability, metabolic effects, and safety. Ann. Rev Nutr. 2002; 22: 19-34.
- **56.** Middleton E. Jr, Kandaswami C., Theoharides T.C.. The effects of plant flavonoids on mammalian cells: implication for inflammation, heart disease and cancer. Pharmacol. Rev. 2000; 52: 673-751.
- **57.** Middleton E Jr., Effect of plant flavonoids on immune and inflammatory cell function. Adv. Exp. Med. Biol. 1998; 439: 175-82.

A Monthly Double-Blind Peer Reviewed Refereed Open Access International e-Journal - Included in the International Serial Directories **International Journal of Research in Medical and Basic Sciences (IJRMS)** http://www.mbsresearch.com email id- irjmss@gmail.com Page 70

- **58.** Zhang B., Safa R. Rusciano D. et al.. Epigallocatechin gallate, an active ingredient from green tea, attenuates damaging influences to the retina caused by ischemia/reperfusion. Brain Res. 2007: 1159: 40-53.
- **59.** Maher P., Hanneken A.. Flavonoids protect retinal ganglion cell from ischemia in vitro. Exp. Eye Res. 2008; 86: 366-74.
- **60.** Maher P., Hanneken A.. Flavonoids protect retinal ganglion cells from oxidative stress-induced death. Invest. Ophthalmol. Vis. Sci. 2005; 46: 4796-803.
- **61.** Jungh S.H., Kang K.D., Ji D. et al.. The flavonoid baicalin counteracts ischemic and oxidative insults to retinal cells and lipid peroxidation to brain membranes. Neurochem. Int. 2008; 53: 325- 37.
- **62.** Heim K.E., Tagliaferro A.R., Bobilya D.J.. Flavonoid antioxidants: chemistry, metabolism and structure-acivity relationships. J. Nutr. Biochem. 2002; 13: 572-84.
- **63.** Ishige K., Schubert D., Sagara Y.. Flavonoids protect neuronal cells from oxidative stress by three distinct mechanisms. Free Radic. Biol. Med. 2001; 30: 433-66.
- **64.** Li G.Y., Fan B., Ma T.H.. Visibile light may directly induce nuclear DNA damage triggering the death pathway in RGC-5 cells. Molecular Vision 2011, 17: 3279-89.
- **65.** Van Bergen N.J., Wood J.P., Chidlow P. et al.. Recharacterization of the RGC-5 retinal ganglion cell line. Invest. Ophthalmol. Vis. Sci. 2009; 50: 4267-72.
- **66.** Green D.R., Reed J.C.. Mithocondria and apoptosis. Science 1998; 281: 1309-12.
- 67. Schmidt H.H., Pollock J.S., Nakane M. et al.. Ca²⁺ /calmodulin nitric oxide sinthases. Cell Calcium 1992; 13: 427-34.
- **68.** Osborne N.N.. Mithocondria: their role in retinal ganglion cell death and survival in primary open-angle glaucoma. Exp. Eye Res. 2010; 90: 750-7.
- **69.** Osborne N.N.. Pathogenesis of ganglion "cell death" in glaucoma and neuroprotection: focus on ganglion cell axonal mithocondria. Progr. Brain Res. 2008; 173: 339-52.
- **70.** Wang Y.S., Xu L., Ma K.. The protective effects of ginkgo biloba extract on cultured human retinal ganglion cells. Zhonghua Yan Ke Za Zhi 2011; 47: 824-8.
- **71.** Fu Q.L., Liao X.X., Li X. et al.. Solubile Nogo-66 receptor prevents synaptic dysfunction and rescues retinal ganglion cells loss in chronic glaucoma. Invest. Ophthalmol. Vis. Sci. 2011; 52: 8374-80.
- **72.** Liao X.X., Chen D., Shi J. et al.. The expression patterns of Nogo-a, myelin-associated glycoprotein and oligodendrocyte myelin glycoprotein in the retina after ocular hypertension: the expression of the myelin proteins in the retina in glaucoma. Neurochem. Res. 2011; 36: 1955-61.

A Monthly Double-Blind Peer Reviewed Refereed Open Access International e-Journal - Included in the International Serial Directories **International Journal of Research in Medical and Basic Sciences (IJRMS)** http://www.mbsresearch.com email id- irjmss@gmail.com Page 71

- **73.** Sippl C., Bosserhoff A.K., Fischer D. et al.. Depletion of optineurin in RGC-5 cells derived from retinal neurons causes apoptosis and reduces the secretion of neurotrophins. Exp. Eye Res. 2011; 93: 669-80.
- **74.** Mori H., Mishina M.. Review: Neurotransmitter receptor VIII: structure and function of the NMDA receptor channel. Neuropharmacology 1995; 34: 1219-37.
- **75.** Chen J., Miao Y., Wang X.H. et al.. Elevation of p-NR2A(S1232) by Cdk5/p35 contributes to retinal ganglion cell apoptosis in a rat experimental glaucoma model. Neurobiol. Dis. 2011; 43: 455-64.
- **76.** Bull N.D., Johnson T.V., Welsapar G. et al.. Use of an adult rat retinal explant model for screening of potential retinal ganglion cell neuroprotective therapies. IOVS 2011 May 17; 52 (6): 3309-20.
- **77.** Yamagishi R., Aihara M., Araie M.. Neuroprotective effects of prostaglandin analogues on retinal ganglion cell death independent of intraocular pressare reduction. Exp. Eye Res. 2011; 93: 265- 70.
- **78.** Cavallotti C., Artico S., Pescosolido N..Age-related changes in the human retina. Can. J. Ophthalmol. 2004: 39: 61-8.
- **79.** Chierzi S., Ratto G.M., Verma P. et al.. The ability of axons to regenerate their growth cones depends on axonal type and age, and is regulated by calcium, cAMP and ERK. Eur. J. Neurosci. 2005; 21: 2051-62.